

# Hepatoprotective Activity of Extracts and Chemically Defined Molecules from Herbal Drugs : Review#

<sup>1</sup>Manoj Kumar Pandey,  
<sup>2</sup>Nitin Rai  
and  
<sup>2</sup>Rajeev Kr. Sharma

<sup>1</sup>Pharmacopoeia Commission for  
Indian Medicine, Raj Nagar,  
Ghaziabad-201002

<sup>2</sup>Pharmacopoeial Laboratory for  
Indian Medicine, Kamla Nehru Nagar,  
Ghaziabad-201002

## Abstract

Liver diseases have become one of the major causes of morbidity and mortality in man and animals all over the globe and hepatotoxicity due to drugs appears to be the most common contributing factor. About 20,000 deaths occur every year due to liver disorders. Hepatocellular carcinoma is one of the ten most common tumors in the world with over 2, 50,000 new cases each year. Plants have potent biochemical components of phytomedicine. Plant based natural phyto-constituents can be derived from any morphological part of the plant and may contain active components. The beneficial medicinal effects of plant materials typically result from the combinations of secondary products present in the plant. The medicinal actions of plants are unique to a particular plant species or groups and are consistent with this concept as the combination of secondary products in a particular plant is taxonomically distinct as such different types of drugs such as acetaminophen; chloroquine and isoniazid are inducing hepatotoxicity in the world. Herbal plants have been used traditionally by herbalists worldwide for the prevention and treatment of liver disease so. Herbal drugs were prescribed even when their biologically active compounds were unknown because of their effectiveness, few side effects and relatively low cost.

In this review, an attempt has been made to compile the reported hepatoprotective activity of plants from India and abroad and may be useful to develop evidence based medicine to cure different kind of liver diseases in man and animal.

**Keywords:** Hepatic disorder, Hepatoprotective herbs, Ayurvedic formulations

## Introduction

Liver has a pivotal role in regulation of physiological processes. It is involved in several vital functions such as metabolism, secretion and storage. Furthermore, detoxification of a variety of drugs and xenobiotics occurs in liver. The bile secreted by the liver has, among other things, an important role in digestion. Liver diseases are among the most serious ailment. They may be classified as acute or chronic hepatitis (inflammatory liver diseases), hepatosis (non inflammatory diseases) and cirrhosis (degenerative disorder resulting in fibrosis of the liver). Liver diseases are mainly caused by toxic chemicals (certain antibiotics, chemotherapeutics, peroxides oil, aflatoxin, carbon-tetrachloride, chlorinated hydrocarbons, etc.), excess consumption of alcohol, infections and autoimmune/disorder.

#Invited Paper

<sup>1</sup>\*Author for correspondence

The liver functions as a great metabolic factory and is particularly concerned with metabolizing drugs, especially those given orally. It plays a key role in the metabolism of lipids, proteins and carbohydrates, as well as in immunomodulation. The sheer complexity and varied nature of its interactions continually expose it to a variety of toxins, therapeutic agents etc., making it susceptible to literally hundreds of diseases. Some of these diseases are rare; others are common, such as hepatitis, cirrhosis, pediatric liver disorders, alcohol related disorders, liver cancer, and weakened liver function on older people. Cirrhosis is the third leading cause of death in adults aged between 25 and 59, and seventh leading cause of death overall. It has been estimated that approximately 14 - 16 million people are infected with this virus in South East Asia region and about 6% of the total population in the region are carriers of this virus.

The situation in India is more serious so far as viral hepatitis is concerned. It is reported that one among every 25 Indians is a carrier of hepatitis B virus and fifth major cause of mortality of people in the age groups of 15-45 years. 1% of total adult death attributed in India is due to infection caused by hepatitis virus B. Besides, incidence 60% chronic liver diseases and 80% of primary liver cancer are due to residual effects of hepatitis B infection. With of lack of safe and effective treatment for liver diseases, researches have been turned towards alternative therapies with ethnic drugs of herbal origin used traditionally, especially in lights of new findings.

Treatment of liver diseases is still largely influenced by holistic approach in different system of medicines. The modern allopathic has very limited effective remedies. However much of remedies claims to be available in folk lore traditional system of medicine based on plants. More than 600 numbers of plants based commercial products are available in different parts of world market for the treatment of variety of liver diseases. In India alone there are more than 60 poly herbal preparations available in market.

*In vivo* and *in vitro* investigations established conclusively that many such plants species does posses prophylactic and therapeutic activity. Recent development of both *in vivo* and *in vitro* investigation procedure laid the foundation for scientific exploration of such plants species as well as helped in validating the folk lore claims. This is evident from the voluminous scientific publications on such investigations on traditional herbal remedies particularly in last two decades. Isolation of novel active phytoconstituents from many such plant species possessing significant potency of antihepatotoxic, will lead for further development of ideal remedies for various liver diseases.

Hence, there is an ever-increasing need for safe hepatoprotective agents. Herbal-based therapeutics for liver disorders have been in use in India for a

long time and popularized world over by leading pharmaceuticals. Despite the significant popularity of several herbal medicines in general, and for liver diseases in particular, they are still unacceptable as treatment modalities for liver diseases. The limiting factors that contribute to this eventuality are lack of standardization of the herbal drugs, identification of active ingredient(s)/principles(s), randomized controlled clinical trials (RCTs) and toxicological evaluation.

A large number of plants and formulations have been claimed to have hepatoprotective activity. Nearly 160 phytoconstituents from 101 plants have been claimed to possess liver protecting activity. In India, more than 87 plants are used in 33 patented and proprietary multi ingredient plant formulations. About 600 commercial herbal formulations with claimed hepatoprotective activity are being sold all over the world. However, only a small proportion of hepatoprotective plants as well as formulations used in traditional medicine are pharmacologically evaluated for their safety and efficacy

Numerous plants and polyherbal formulations are used for the treatment of liver diseases. However, in most of the severe cases, the treatments are not satisfactory. Although experimental evaluations were carried out on a good number of these plants and formulations, the studies were mostly incomplete and insufficient. The therapeutic values were tested against a few chemicals-induced subclinical levels of liver damages in rodents. Even common dietary antioxidants can provide such protection from liver damage caused by oxidative mechanisms of toxic chemicals. However, experiments have clearly shown that plants such as *Picrorrhiza kurroa*, *Andrographis paniculata*, *Eclipta alba*, *Silibum marianum*, *Phyllanthus maderaspatensis* and *Trichopus zeylanicus* are sufficiently active against, at least, certain hepatotoxins.

Single plant may not have all the desired activities. A combination of different herbal extracts/fractions is likely to provide desired activities to cure severe liver diseases. Development of such medicines with standards of safety and efficacy can revitalise treatment of liver disorders and hepatoprotective activity.

The traditional medicinal plants species have been subjected in various experimental models of investigation and attempt has made to calibrate their therapeutical activity. The herbs used in hepatic disease have been extensively exploited all over the world and large numbers of the plant species has been documented as hepatoprotective antihepatotoxic, cholegoge and choleric. A diverse nature of chemical compounds has been identified from such plants species through bio-assay guided investigation. However, in some plant species the total extract or fraction of extract has reported to possess better and potent biological activity compared to isolated pure compound(s) from the

extracts. Plant derived extracts and chemically defined molecules are enumerated in Table 1 and 2.

**Table 1:** Plant Extracts with Hepatoprotective Activity

Family and Botanical name	Origin	Plant Parts used	Extracts Studied	Type of assay	Hepato-toxicity Inducing Agents	References
Acanthaceae <i>Acanthus ilicifolius</i> L.	India	Leaves	Alcohol	<i>In vivo</i>	CCl <sub>4</sub>	Babu <i>et al.</i> (2001)
<i>Andrographis lineata</i> Nees	India	Leaves	Aqueous, methanol	<i>In vivo</i>	CCl <sub>4</sub>	Sangameswaran <i>et al.</i> (2008)
<i>Andrographis paniculata</i> (Burm.f.) Nees	India	Leaves	Alcohol	<i>In vivo</i>	CCl <sub>4</sub>	Rana and Avadhoot (1991)
<i>Anisotes trisulcus</i> (Forssk.)	Yemen	Whole plant	Ethanol	<i>In vivo</i>	CCl <sub>4</sub>	Fleurentin <i>et al.</i> (1986)
<i>Asteracantha longifolia</i> L.	Sri Lanka	Whole plant	Aqueous	<i>In vivo</i>	CCl <sub>4</sub> and PCM	Hewawasam <i>et al.</i> (2003)
<i>Hygrophila auriculata</i> (K.Schum.) Heine	India	Seeds	Methanol	<i>In vivo</i>	PCM and Thioacetamide	Singh and Handa (1995)
<i>Hypoestes triflora</i> (Forssk.) Roem. and Schult	Rwanda	Leaves	Aqueous	<i>In vivo</i>	CCl <sub>4</sub>	Van Puyvelde <i>et al.</i> (1989)
<i>Rhinacanthus nasuta</i> (L.) Kurz.	India	Root	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Suja <i>et al.</i> (2003)
Adoxaceae <i>Viburnum tinus</i> L.	Southern Europe	Leaves	Aqueous-methanol	<i>In vivo</i>	CCl <sub>4</sub>	Mohammed <i>et al.</i> (2005)
Aizoaceae <i>Trianthema portulacastrum</i> L.	India	Leaves	Ethanol	<i>In vivo</i>	PCM and Thioacetamide	Kumar <i>et al.</i> (2004)
Apiaceae <i>Apium graveolens</i> L.	India	Seeds	Methanol	<i>In vivo</i>	PCM and Thioacetamide	Singh and Handa (1995)
<i>Carum copticum</i> L.	Pakistan	Seeds	Aqueous-methanol	<i>In vivo</i>	CCl <sub>4</sub> and PCM	Gilani <i>et al.</i> (2005a)
Apocynaceae <i>Apocynum venetum</i> L.	China, Japan	Leaf	Aqueous	<i>In vivo</i>	CCl <sub>4</sub> and GAIN	Xiong <i>et al.</i> (2000)
Araliaceae <i>Acanthopanax senticosus</i> (Rupr. and Maxim.) Harms	Taiwan	Whole plant	Methanol	<i>In vivo</i>	CCl <sub>4</sub> and Acetaminophen	Lin and Huang (2002)
Asclepiadaceae <i>Sarcostemma brevistigma</i> Wight	India	Stem bark	Ethyl acetate	<i>In vivo</i>	CCl <sub>4</sub>	Sethuraman <i>et al.</i> (2003)
Asteraceae <i>Achyrocline satureioides</i> (Lam.) DC.	Argentina	Aerial parts	Aqueous	<i>In vivo</i>	Bromobenzene	Kadarian <i>et al.</i> (2002)

Family and Botanical name	Origin	Plant Parts used	Extracts Studied	Type of assay	Hepato-toxicity Inducing Agents	References
<i>Artemisia absinthium</i> L.	Pakistan	Aerial parts	Aqueous-methanol	<i>In vivo</i>	CCl <sub>4</sub> and Aceta-minophen	Gilani and Janbaz (1995a)
<i>Artemisia maritima</i> L.	Pakistan	Aerial parts	Aqueous-methanol	<i>In vivo</i>	CCl <sub>4</sub> and Aceta-minophen	Janbaz and Gilani (1995)
<i>Artemisia vulgaris</i> L.	Pakistan	Aerial parts	Aqueous-methanol	<i>In vivo</i>	GAIN and LPS	Gilani <i>et al.</i> (2005b)
<i>Bidens chilensis</i> DC	Taiwan	Whole plant	Methanol	<i>In vivo</i>	CCl <sub>4</sub> and PCM	Chih <i>et al.</i> (1996)
<i>Bidens pilosa</i> L.	Taiwan	Whole plant	Methanol	<i>In vivo</i>	CCl <sub>4</sub> and PCM	Chih <i>et al.</i> (1996)
<i>Cichorium intybus</i> L.	India	Seeds	Alcohol	<i>In vivo</i>	CCl <sub>4</sub>	Ahmed <i>et al.</i> (2003)
<i>Crassocephalum crepidioides</i> Benth	Japan	Whole plant	Aqueous	<i>In vivo</i> and <i>in vitro</i>	GAIN, LPS and CCl <sub>4</sub>	Aniya <i>et al.</i> (2005)
<i>Elephantopus mollis</i> Kunth.	Taiwan	Whole plant	Aqueous	<i>In vivo</i>	Acetaminophen and GAIN	Lin <i>et al.</i> (1995b)
<i>Elephantopus scaber</i> L.	Taiwan	Whole plant	Aqueous	<i>In vivo</i>	Acetaminophen and GAIN	Lin <i>et al.</i> (1995b)
<i>Flaveria trinervia</i> (Spreng.) C.Mohr	India	Leaf	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Umadevi <i>et al.</i> (2004)
<i>Gundelia tourenfortii</i> L.	Iran	Stalk	Hydro-alcoholic	<i>In vivo</i> and <i>in vitro</i>	CCl <sub>4</sub>	Jamshidzadeh <i>et al.</i> (2005)
<i>Pseudelephantopus spicatus</i> (Juss. Ex Aublet) Gleason	Taiwan	Whole plant	Aqueous	<i>In vivo</i>	Acetaminophen and GAIN	Lin <i>et al.</i> (1995b)
<i>Wedelia chinensis</i> (Osbeck) Merr.	Taiwan	Whole plant	Methanol	<i>In vivo</i>	CCl <sub>4</sub> , Aceta-minophen and GAIN	Lin <i>et al.</i> (1994)
<i>Wedelia calendulacea</i> L.	India	Leaf	Ethanol	<i>In vivo</i>	CCl <sub>4</sub>	Murugaian <i>et al.</i> (2008)
Balanophoraceae <i>Thonningia sanguinea</i> Vahl.	Ghana	Roots, leaves	Aqueous	<i>In vivo</i> and <i>in vitro</i>	GAIN and CCl <sub>4</sub>	Gyamfi <i>et al.</i> (1999)
Bixaceae <i>Cochlospermum tinctorium</i> Perri ex Rich.	Mali	Rhizome	Ethanol and hydro-ethanol extract	<i>In vivo</i>	CCl <sub>4</sub>	Diallo <i>et al.</i> (1992)
<i>Bixa orellana</i> L.	Bangladesh	Seed	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Ahsan <i>et al.</i> (2009)
Brassicaceae <i>Coronopus didymus</i> L.	India	Whole plant	Aqueous	<i>In vivo</i>	CCl <sub>4</sub>	Mantena <i>et al.</i> (2005)

Family and Botanical name	Origin	Plant Parts used	Extracts Studied	Type of assay	Hepato-toxicity Inducing Agents	References
Burseraceae <i>Commiphora opobalsamum</i> (L.) Engl.	Saudi Arabia	Aerial parts	Ethanol	<i>In vivo</i>	CCl <sub>4</sub>	Al-Howiriny <i>et al.</i> (2004)
Caesalpiniaceae <i>Bauhinia racemosa</i> Lam	India	Bark	Methanol	<i>In vivo</i>	CCl <sub>4</sub> and PCM	Gupta <i>et al.</i> (2004)
Capparidaceae <i>Cleome viscosa</i> L.	India	Leaves	Ethanol	<i>In vivo</i>	CCl <sub>4</sub>	Gupta <i>et al.</i> (2009)
Casuarinaceae <i>Casuarina equisetifolia</i> Forst	Bangladesh	Leaves, bark	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Ahsan <i>et al.</i> (2009)
Celasteraceae <i>Salacia reticulata</i> Wight	Sri Lanka, India	Root, stem	Aqueous, methanol	<i>In vivo</i>	CCl <sub>4</sub>	Yoshikawa <i>et al.</i> (2002)
Chenopodiaceae <i>Beta vulgaris</i> L.	India	Root	Ethanol	<i>In vivo</i>	CCl <sub>4</sub>	Agarwal <i>et al.</i> (2006)
Combretaceae <i>Combretum</i> Kurz.	Japan	Leaves	Methanol	<i>In vivo</i> and <i>in vitro</i>	GAIN	Banskota <i>et al.</i> (2003)
<i>Terminalia arjuna</i> L.	India	Bark	Aqueous	<i>In vivo</i>	CCl <sub>4</sub>	Manna <i>et al.</i> (2006)
<i>Terminalia belerica</i> Roxb	India	Fruits	Ethanol	<i>In vivo</i>	CCl <sub>4</sub>	Jadon <i>et al.</i> (2007)
<i>Terminalia catappa</i> L.	Okinawa Island	Leaves	Aqueous	<i>In vivo</i> and <i>in vitro</i>	GAIN and LPS	Kinoshita <i>et al.</i> (2007)
<i>Terminalia chebula</i> Reiz.	India	Fruits	Ethanol	<i>In vivo</i> and <i>in vitro</i>	Anti TB drugs	Tasduq <i>et al.</i> (2006)
Compositae <i>Ambrosia maritima</i> L.	Egypt	Whole plant	Aqueous-methanol	<i>In vivo</i>	Acetaminophen	Ahmed and Kharter (2001)
<i>Crepis rueppellii</i> (Sch.) Bip.	Yemen	Whole plant	Ethanol	<i>In vivo</i>	CCl <sub>4</sub>	Fleurentin <i>et al.</i> (1986)
<i>Eclipta alba</i> Hassk.	India	Whole plant	Alcohol	<i>In vivo</i>	CCl <sub>4</sub>	Singh <i>et al.</i> (1993)
<i>Epaltes divaricata</i> (L.) Cav.	India	Whole plant	Aqueous	<i>In vivo</i>	CCl <sub>4</sub>	Hewawasam <i>et al.</i> (2004)
Convolvulaceae <i>Cuscutae semen</i> Lam.	Korea	Seeds	Aqueous	<i>In vivo</i>	DMN	Kim <i>et al.</i> (2007a)
<i>Erycibe expansa</i> Wall. and G.Don	Thailand	Stem	Methanol	<i>In vitro</i>	GAIN	Matsuda <i>et al.</i> (2004)
Crassulaceae <i>Kalanchoe pinnata</i> Pers.	India	Leaves	Juice of leaves, ethanol extract of marc	<i>In vivo</i> and <i>in vitro</i>	CCl <sub>4</sub>	Yadav and Dixit (2003)

Family and Botanical name	Origin	Plant Parts used	Extracts Studied	Type of assay	Hepato-toxicity Inducing Agents	References
Cucurbitaceae <i>Luffa echinata</i> Roxb.	India	Fruits	Pet.ether, acetone, methanol	<i>In vivo</i>	CCl <sub>4</sub>	Ahmed <i>et al.</i> (2002)
Cyperaceae <i>Cyperos scariosus</i> R.Br.	Indonesia, Pakistan	Tubers	Aqueous-methanol	<i>In vivo</i>	CCl <sub>4</sub>	Gilani and Jambaz (1995b)
Ebenaceae <i>Diospyros malabarica</i> (Desr.) Kostel	India	Bark	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Mondal <i>et al.</i> (2005)
Euphorbiaceae <i>Alchornea cordifolia</i> Schum and Thonn.	Nigeria	Leaves	Ethanol	<i>In vivo</i>	Acetaminophen	Olaleye <i>et al.</i> (2006)
<i>Croton oblongifolius</i> Roxb.	India	Aerial parts	Pet.ether, acetone, methanol	<i>In vivo</i>	CCl <sub>4</sub>	Ahmed <i>et al.</i> (2002)
<i>Embllica officinalis</i> Gaertner	India	Fruits	Hydro-alcoholic	<i>In vitro</i>	Anti TB drugs	Tasduq <i>et al.</i> (2005)
<i>Phyllanthus maderaspatensis</i> L.	India	Whole plant	n-hexane	<i>In vivo</i>	CCl <sub>4</sub> and Thioacetamide	Asha <i>et al.</i> (2007)
<i>Phyllanthus niruri</i> L.	Brazil	Leaves	Aqueous	<i>In vivo</i>	PCM	Sabir and Rocha (2008)
<i>Phyllanthus reticulatus</i> Poir.	India	Aerial parts		Ethanol	<i>In vivo</i>	CCl <sub>4</sub> Das <i>et al.</i> (2008)
Fabaceae <i>Acacia catechu</i> (L.f.) Willd.	India	Bark	Ethyl acetate		<i>In vivo</i>	CCl <sub>4</sub> Ray <i>et al.</i> (2006)
<i>Bauhinia variegata</i> L. bark	India	Stem	Alcohol	<i>In vivo</i>	CCl <sub>4</sub>	Bodakhe and Ram (2007)
<i>Cajanus cajan</i> L.	India	Leaves	Methanol	<i>In vivo</i>	Alcohol	Kundu <i>et al.</i> (2008)
<i>Cassia fistula</i> L.	India	Leaves	n-heptane	<i>In vivo</i>	CCl <sub>4</sub>	Bhakta <i>et al.</i> (1999)
<i>Cassia occidentalis</i> L.	India	Leaves	Aqueous-ethanol	<i>In vivo</i>	PCM and ethyl alcohol	Jafri <i>et al.</i> (1999)
<i>Glycine max</i> (L.) Merr	Taiwan	Seed	Water	<i>In vivo</i>	Acetaminophen	Wu <i>et al.</i> (2001)
<i>Phaseolus aureus</i> Roxb.	Taiwan	Seed	Water	<i>In vivo</i>	Acetaminophen	Wu <i>et al.</i> (2001)
<i>Phaseolus calcaratus</i> Roxb	Taiwan	Seed	Water	<i>In vivo</i>	Acetaminophen	Wu <i>et al.</i> (2001)
<i>Phaseolus radiatus</i> L.	Taiwan	Seed	Water	<i>In vivo</i>	Acetaminophen	Wu <i>et al.</i> (2001)
<i>Pterocarpus marsupium</i> Roxb.	India	Stem bark	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Mankani <i>et al.</i> (2005)
<i>Trigonella foenum-graecum</i> L.	India	Leaves	Ethanol	H <sub>2</sub> O <sub>2</sub>	CCl <sub>4</sub>	Meera (2009)

Family and Botanical name	Origin	Plant Parts used	Extracts Studied	Type of assay	Hepato-toxicity Inducing Agents	References
Fumariaceae <i>Fumaria indica</i> (Hauskn.) Pugsley	India	Whole plant	Methanol, Pet.Ether, aqueous	<i>In vivo</i>	PCM, Rifampicin, CCl <sub>4</sub>	Rao and Mishra (1997)
<i>Fumaria parviflora</i> Lam.	Pakistan	Shoots	Aqueous-methanol	<i>In vivo</i>	PCM	Gilani <i>et al.</i> (1996)
Gentianaceae <i>Enicostemma littorale</i> Blume.	India	Whole plant	Alcohol	<i>In vivo</i>	CCl <sub>4</sub>	Senthilkumar <i>et al.</i> (2005)
<i>Swertia japonica</i> (Roem. and Schult.) Makino.	Japan	Whole plant	Butanol	<i>In vivo</i>	GAIN	Hase <i>et al.</i> (1997b)
Lamiaceae <i>Ocimum basilicum</i> L.	India	Leaves	Ethanol	H <sub>2</sub> O <sub>2</sub>	CCl <sub>4</sub>	Meera <i>et al.</i> (2009)
Moraceae <i>Ficus carica</i> L.	India	Leaves	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Krishna <i>et al.</i> (2007)
<i>Ficus hispida</i> L.	India	Leaves	Methanol	<i>In vivo</i>	PCM	Mandal <i>et al.</i> (2000)
Moringaceae <i>Moringa oleifera</i> L.	Malaysia	Leaves	Hydro-alcoholic	<i>In vivo</i>	Acetaminophen	Fakurazi <i>et al.</i> (2008)
Myrtaceae <i>Careya arborea</i> Roxb.	India	Stem bark	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Sambath <i>et al.</i> (2005)
Nyctaginaceae <i>Boerhaavia diffusa</i> L.	India	Roots	Aqueous	<i>In vivo</i>	Thioacetamide	Rawat <i>et al.</i> (1997)
Nymphaceae <i>Nymphaea stellata</i> Willd.	India	Flowers	Alcohol	<i>In vivo</i>	CCl <sub>4</sub>	Bhandarkar and Khan (2004)
Oleaceae <i>Phillyrea latifolia</i> L.	Jordan	Leaves	Aqueous	<i>In vivo</i>	CCl <sub>4</sub>	Janakat and Al-Merie (2002)
Ophioglossaceae <i>Helminthostachys zeylanica</i> (L.) Hook	India	Rhizomes	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Suja <i>et al.</i> (2004)
Orchidaceae <i>Anoectochilus formosanus</i> Hayata	Taiwan	Whole plant	Aqueous	<i>In vivo</i> and <i>in vitro</i>	CCl <sub>4</sub>	Wu <i>et al.</i> (2007)
Polygalaceae <i>Polygala arvensis</i> Willd.	India	Leaves	Chloroform	<i>In vivo</i>	GAIN	Dhanabal <i>et al.</i> (2006)
Rhamnaceae <i>Ventilago leiocarpa</i> Benth.	Taiwan	Bark	Methanol, ethanol, butanol and aqueous	<i>In vivo</i>	CCl <sub>4</sub>	Lin <i>et al.</i> (1995a)
<i>Ziziphus mauritiana</i> Lam.	Nigeria	Leaves	Ethanol	<i>In vivo</i>	CCl <sub>4</sub>	Dahiru <i>et al.</i> (2005)



Family and Botanical name	Origin	Plant Parts used	Extracts Studied	Type of assay	Hepato-toxicity Inducing Agents	References
Rubiaceae <i>Hedyotis corymbosa</i> (L.) Lam.	India	Whole plant	Methanol	<i>In vivo</i>	PCM	Sadasivan <i>et al.</i> (2006)
<i>Mitracarpus scaber</i> Zucc.	Mali	Whole plant	Methanol	<i>In vivo</i> and <i>in vitro</i>	CCl <sub>4</sub>	Germano <i>et al.</i> (1999)
<i>Morinda citrifolia</i> L.	America	Whole plant	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Wang <i>et al.</i> (2008)
Rutaceae <i>Aegle marmelos</i> (L.) Corr. Serr.	India	Leaves	Fine powder in physiological saline	<i>In vivo</i>	Alcohol	Singanani <i>et al.</i> (2007)
<i>Glycosmis pentaphylla</i> Corr.	Bangladesh	Leaves, bark	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Ahsan <i>et al.</i> (2009)
Scrophulariaceae <i>Bacopa monniera</i> (L.) Pennell	India	Whole plant	Alcohol	<i>In vivo</i>	Morphine	Sumathy <i>et al.</i> (2001)
<i>Picrorrhiza kurroa</i> (Roule.) Sans	Himalayas	Rhizome, roots	Ethanol	<i>In vivo</i>	GAIN	Anandan and Devaki (1999)
Smilacaceae <i>Smilax regelii</i> Killip and Morton	Saudi Arabia	Roots	Ethanol	<i>In vivo</i>	CCl <sub>4</sub>	Rafatullah <i>et al.</i> (1991)
Solanaceae <i>Nicotiana glauca</i> Graham.	Jordan	Leaves, flowers	Aqueous	<i>In vivo</i>	CCl <sub>4</sub>	Janakat and Al-Merie (2002)
<i>Solanum nigrum</i> L.	India	Fruits	Ethanol	<i>In vivo</i>	CCl <sub>4</sub>	Raju <i>et al.</i> (2003)
<i>Solanum pseudocapsicum</i> Hassl.	Jerusalem	Leaves	Methanol	<i>In vivo</i> and <i>in vitro</i>	CCl <sub>4</sub>	Vijayan <i>et al.</i> (2003)
<i>Solanum trilobatum</i> L.	India	Whole plant	Methanol	<i>In vivo</i>	CCl <sub>4</sub>	Shahjahan <i>et al.</i> (2004)
Umbelliferae <i>Bupleurum kaoi</i> Liu (Chao et Chuang)	Taiwan	Leaves	Aqueous	<i>In vitro</i>	Acetaminophen and CCl <sub>4</sub>	Liu <i>et al.</i> (2006)
<i>Daucus carota</i> L.	Europe, Asia, Africa	Roots	Aqueous	<i>In vivo</i>	CCl <sub>4</sub>	Bishayee <i>et al.</i> (1995)
<i>Foeniculum vulgare</i>	Turkey Miller	Seeds	Essential oil	<i>In vivo</i>	CCl <sub>4</sub>	Ozbek <i>et al.</i> (2003)
Valerianaceae <i>Nardostachys jatamansi</i> D.C.	India	Rhizomes	Ethanol	<i>In vivo</i>	CCl <sub>4</sub>	Ali <i>et al.</i> (2000)
Vitaceae <i>Rhoicissus tridentata</i> (L.f.) Wild and R.B. Drumm	South Africa	Roots	Aqueous	<i>In vivo</i>	CCl <sub>4</sub>	Opoku <i>et al.</i> (2007)

**Table 2:** Chemically defined Molecules with Hepatoprotective activity

Chemical substance	Plant	Plant part	Class	References
3,4-di-O-caffeoylquinic acid	<i>Lactuca indica</i> L.	Aerial parts	Quinic acid	Kim <i>et al.</i> (2007b)
3,5-di-O-caffeoyl-muco-quinic acid	<i>Lactuca indica</i> L.	Aerial parts	Quinic acid	Kim <i>et al.</i> (2007b)
5-O-(E)-p-coumaroylquinic acid	<i>Lactuca indica</i> L.	Aerial parts (2007b)	Quinic acid	Kim <i>et al.</i>
$\alpha$ -Amyrin (Aubl.) March	<i>Protium heptaphyllum</i>	Trunk wood resin	Triterpene	Oliveira <i>et al.</i> (2005)
$\beta$ -Amyrin (Aubl.) March	<i>Protium heptaphyllum</i>	Trunk wood resin	Triterpene	Oliveira <i>et al.</i> (2005)
Anastatin A	<i>Anastatica hierochuntica</i> L.	Whole plant	Flavonoid	Yoshikawa <i>et al.</i> (2003)
Anastatin B	<i>Anastatica hierochuntica</i> L.	Whole plant	Flavonoid	Yoshikawa <i>et al.</i> (2003)
18 $\beta$ -glycyrrhetic acid	<i>Glycyrrhiza uralensis</i> Fisch.	Rhizomes	Glycyrrhetic acid	Shim <i>et al.</i> (2000)
Tetrahydroswertianolin	<i>Swertia japonica</i> Makino	Aerial parts	Xanthione	Hase <i>et al.</i> (1997b)
Gentiopicroside	<i>Swertia japonica</i> Makino	Aerial parts	Iridoid	Hase <i>et al.</i> (1997b)
Sweroside	<i>Swertia japonica</i> Makino	Aerial parts	Iridoid	Hase <i>et al.</i> (1997b)
Andrographolide	<i>Andrographis paniculata</i> (Burm.f) Nees	Aerial parts	Diterpene	Chander <i>et al.</i> (1995)
Erycibenin A	<i>Erycibe expansa</i>	Stem	Pterocarpane	Matsuda <i>et al.</i> (2004)
5,7,4'-trihydroxy-3'-Methoxyisoflavone	<i>Erycibe expansa</i>	Stem	Isoflavone	Matsuda <i>et al.</i> (2004)
Genistein	<i>Erycibe expansa</i> Wall. Ex G. Don.	Stem	Isoflavone	Matsuda <i>et al.</i> (2004)
Orobol	<i>Erycibe expansa</i> Wall. Ex G. Don.	Stem	Isoflavone	Matsuda <i>et al.</i> (2004)
Mangiferin	<i>Salacia reticulata</i> Abst.	Roots	Phenolic compound	Yoshikawa <i>et al.</i> (2002)
(-)-4'-O-methyl-epigallocatechin	<i>Salacia reticulata</i> Abst.	Roots	Phenolic compound	Yoshikawa <i>et al.</i> (2002)
Thymoquinone	<i>Nigella sativa</i> L.	Aerial parts	Quinone	Daba and Abdel-Rahman (1998)
Lithospermate B	<i>Salvia miltorhiza</i> Bunge	Roots	Caffeic acid	Hase <i>et al.</i> (1997a)

Chemical substance	Plant	Plant part	Class	References
Taxiresinol	<i>Enciostemma littorale</i>	Aerial parts	Tetrahydrofuran	Nguyen <i>et al.</i> (2004)
(7'R)-7'-hydroxylariciresinol	<i>Enciostemma littorale</i>	Aerial parts	Tetrahydrofuran	Nguyen <i>et al.</i> (2004)
Onitin	<i>Equisetum arvense</i> L.	Aerial parts	Phenolic compound	Oh <i>et al.</i> (2004)
Luteolin	<i>Equisetum arvense</i> L.	Aerial parts	Flavonoid	Oh <i>et al.</i> (2004)
Quercetin-3-O-β-D-glucuronopyranoside	<i>Saururus chinensis</i> (Lour.) Baill.	Aerial parts	Flavonol glycoside	Sung <i>et al.</i> (1997)
Quercetin-3-O-β-D-glucuronopyranosyl methyl ester	<i>Saururus chinensis</i> (Lour.) Baill.	Aerial parts	Flavonol glycoside	Sung <i>et al.</i> (1997)
Scropolioside-A	<i>Scrophularia koelzii</i> Pennell	Aerial parts	Iridoid glycoside	Garg <i>et al.</i> (1994)
3-(S)-3-_-Dglucopyranosyloxybutanolide	<i>Goodyera schlechtendaliana</i> Reichb.G. <i>matsumurana</i> Schltr.G. <i>discolor</i> Kergawl	Whole plant	Aliphatic glycoside	Du <i>et al.</i> (2000)
3-(S)-3-_-D-glucopyranosyloxy-4-hydroxybutanoic acid	<i>Goodyera schlechtendaliana</i> Reichb.G. <i>matsumurana</i> Schltr.G. <i>discolor</i> Kergawl.	Whole plant	Aliphatic glycoside	Du <i>et al.</i> (2000)
Agathisflavone	<i>Canarium manii</i> King	Aerial parts	Biflavonoid	Anand <i>et al.</i> (1992)
(S)-bakuchiol	<i>Psoralea corylifolia</i> Babchi	Aerial parts	Monoterpene phenol	Hyun <i>et al.</i> (2001)
Monomethyl fumarate	<i>Fumaria indica</i> Pugsley	Whole plant	Fumaric acid	Rao and Mishra (1998)
Wighteone	<i>Cudrania cochinchinensis</i> (Lour.) Kudo et Masam.	Roots	Flavonoid	Lin <i>et al.</i> (1996)
Naringenin	<i>Cudrania cochinchinensis</i> (Lour.) Kudo et Masam.	Roots	Flavonoid	Lin <i>et al.</i> (1996)
Torilin	<i>Cnidium monnieri</i> (L.) Cusson	Aerial parts	Sesquiterpene	Oh <i>et al.</i> (2002)
Torilolone	<i>Cnidium monnieri</i> (L.) Cusson.	Aerial parts	Sesquiterpene	Oh <i>et al.</i> (2002)
Allicin	<i>Allium sativum</i> L.	Cloves	Allyl thiosulfates	Vimal and Devaki (2004)
Kaempferol	<i>Rhodiola sachalinensis</i> A.Bor.	Roots	Phenolic compound	Song <i>et al.</i> (2003)

Chemical substance	Plant	Plant part	Class	References
Salidroside	<i>Rhodiola sachalinensis</i> A.Bor.	Roots	Phenolic compound	Song <i>et al.</i> (2003)
1-O-galloyl-6-O-(4-hydroxy-3,5-dimethoxy)benzoyl- $\beta$ -D-glucose	<i>Combretum quadrangulare</i> Kurz	Seeds	Gallic acid	Adnyana <i>et al.</i> (2001)
Picroliv	<i>Picrorhiza kurroa</i> Royle ex Benth.	Aerial parts	Iridoid glycoside	Visen <i>et al.</i> (1991)
Indigtone	<i>Indigofera tinctoria</i> L.	Aerial parts	Aliphatic nitrocompound	Singh <i>et al.</i> (2001)
Acanthoic acid	<i>Acanthopanax koreanum</i> Nakai	Root bark	Diterpene	Park <i>et al.</i> (2004)
Myristin	<i>Myristica fragrans</i> Houtt.	Aerial parts	Cetyl ester	Morita <i>et al.</i> (2003)
Rutin	<i>Artemisia scoparia</i> Waldst. and Kit.	Aerial parts	Flavonoid	Janbaz <i>et al.</i> (2002)
Troxeutin	<i>Artemisia scoparia</i> Waldst. and Kit.	Aerial parts	Flavonoid	Zhang <i>et al.</i> (2009)
Neoandrographolide	<i>Andrographis paniculata</i> (Burm.f.) Wall. Ex Nees	Aerial parts	Diterpene	Chander <i>et al.</i> (1995)
5-O-methyl-(E)-resveratrol.3-O- $\beta$ -D-glucopyranoside	<i>Acer mono</i> Maxim.	Leaves	Stilbene glycoside	Yang <i>et al.</i> (2005)
5-O-methyl-(E)-resveratrol.3-O- $\beta$ -D-apiofuranosyl-1- $\rightarrow$ 6)- $\beta$ -D-glucopyranoside	<i>Acer mono</i> Maxim.	Leaves	Stilbene glycoside	Yang <i>et al.</i> (2005)
Corilagin	<i>Terminalia catappa</i> L.	Leaves	Tannin	Kinoshita <i>et al.</i> (2007)
$\gamma$ -Amyrone	<i>Sedum sarmentosum</i> Bunge	Aerial parts	Triterpene	Amin <i>et al.</i> (1998)
3-epi- $\gamma$ -amyrin	<i>Sedum sarmentosum</i> Bunge	Aerial parts	Triterpene	Amin <i>et al.</i> (1998)
$\gamma$ -Amyrin	<i>Sedum sarmentosum</i> Bunge	Aerial parts	Triterpene	Amin <i>et al.</i> (1998)
18 $\beta$ -hydroperoxy-olean.12-en-3-one	<i>Sedum sarmentosum</i> Bunge	Aerial parts	Triterpene	Amin <i>et al.</i> (1998)
Rubiadin	<i>Rubia cordifolia</i> L.	Roots	Anthraquinone	Rao <i>et al.</i> (2006)
3,4,5-trihydroxybenzoic acid	<i>Terminalia bellerica</i> Roxb.	Fruit	Gallic acid	Jadon <i>et al.</i> (2007)
Kinsenoside	<i>Anoectochilus formosanus</i> Hay.	Whole plant	Furanone	Wu <i>et al.</i> (2007)

## Discussion and Conclusion

The liver is the most important organ in the body. It has a pivotal role in regulation of physiological processes. It is involved in several vital functions such as metabolism, secretion and storage. Liver diseases are among the most serious ailments. They may be classified as acute or chronic hepatitis (inflammatory liver diseases), hepatosis (non inflammatory diseases) and cirrhosis (degenerative disorder resulting in fibrosis of the liver).

Modern society has innate knowledge about the herbal treatment of liver disease from many cultures. Research into plants traditionally used in the treatment of liver disease has significantly advanced in the past 15 years, and much of what has been discovered supports traditional knowledge. Considering the enormous biodiversity resources over the world's traditional system and the high incidence of liver complications, the present review extensively focuses on collection of data for different plants, which are available in India and all over the world. These medicinal plants claimed as liver protective agents are classified according to their biological source, phytoconstituents; part used and plants in formulations. People from India are still dependent on conventional therapies to treat liver complications. Because of their easy availability and low cost. Since large mass of populations used preferable herbal preparation, therefore there is need to be evaluate for their proportion, their dose and rational behind combination in different polyherbal preparation.

These herbal drugs have shown the ability to maintain the normal functional statuses of the liver with or without fewer side effects. These are the reason that's why herbal hepatoprotectives are mostly preferred by medical practitioners.

It has been seen that herbal hepatoprotective drugs have less side effect or interaction as compared to synthetic medicine but in other hand scientific evidence from tests done to evaluate the safety and effectiveness of traditional hepatoprotective medicine products and practices is limited and further study of products and practices is needed.

Pharmacokinetic and toxicity studies have not disclosed any issues that could limit the therapeutic use of these drugs. Also the study is required to identify glycosides, flavonoids, triterpenes and phenolic compounds as classes of compounds with hepatoprotective activity.

Further studies including clinical trials need to be carried out to ascertain the safety of these compounds as a good alternative to conventional drugs in the treatment of liver diseases

Since the traditional system of medicine recommends various hepatoprotective agents and preparations to treat hepatic disorders. The management of liver disease is still challenges to modern medicine. The modern allopathic drugs have very little to offer for alleviation of hepatic ailments and some these drugs adversely affect the liver function. A phytotherapeutic approach to modern drug development provides many invaluable drugs from traditional medicinal plants. Search for pure phytochemical as drug is time consuming and extensive. Numerous plants and polyherbal formulations are being used for the treatment of liver diseases.

Today, unfortunately the herbal resources have declined rapidly because more than 80% of our total medicinal plants used by Indian pharmaceutical industry are collected from their wild sources and they are not being grown or domesticated so far. To meet their burgeoning demand is the need of the day. Moreover, our natural resource base of medicinal plants is being depleting day by day. Hence there is an urgent need to encourage field scale cultivation of prioritized medicinal plants through government initiatives before it's too late.

## References

- Adnyana, K., Tezuka, Y., Awale, S., Banskota A.H., Kim, Q.T. and Kadota, S., 2001. 1-O-galloyl-6-O-(4-hydroxy-3, 5-dimethoxy) benzoyl- $\beta$ -D-glucose, a new hepatoprotective constituent from *Combretum quadrangulare*. *Planta Med.* 67(4): 370-371.
- Agarwal, M., Srivastava, V.K, Saxena, K.K. and Kumar, A., 2006. Hepatoprotective activity of *Beta vulgaris* against CCl<sub>4</sub>- induced hepatic injury in rats. *Fitoterapia* 77(2): 91-93.
- Ahmed, B., Alam, T., Varshney, M. and Khan, S.A., 2002. Hepatoprotective activity of two plants belonging to the Apiaceae and the Euphorbiaceae family. *J. Ethnopharmacol.* 79(3): 313-316.
- Ahmed, B., Al-Howiriny, T.A. and Siddiqui, A.B., 2003. Antihepatotoxic activity of seeds of *Cichorium intybus*. *J. Ethnopharmacol* 87: 37-240.
- Ahmed, B.M. and Khater, R.M., 2001. Evaluation of the protective potential of *Ambrosia maritima* extract on acetaminophen induced liver damage. *J. Ethnopharmacol.* 75: 169-171.
- Ahsan, M.R., Islam, K.M. and Bulbul, I.J., 2009. Hepatoprotective activity of Methanol Extract of some medicinal plants against carbon tetrachloride-induced hepatotoxicity in rats. *Eur. J. Sci. Res.* 37(2): 302-310.
- Ali, S., Ansari, K.A., Jafry, M.A., Kabeer, H. and Diwakar, G., 2000. *Nardostachys jatamansi* protects against liver damage induced by thioacetamide in rats. *J. Ethnopharmacol.* 71(3): 359-363.

- Al-Howiriny, T.A., Al-Sohaibani, M.O., Al-Said, M.S., Al-Yahya, M.A., El-Tahir, K.H. and Rafatullah, S., 2004. Hepatoprotective properties of *Commiphora opobalsamum* (Balessan), a traditional medicinal plant of Saudi Arabia. *Drugs Exp. Clin. Res.* 30: 213-220.
- Amin, H., Mingshi, W., Hong, Y.H., Decheng, Z. and Lee, K.H., 1998. Hepatoprotective triterpenes from *Sedum sarmentosum*. *Phytochem.* 49(8): 2607-2610.
- Anand, K.K., Gupta, V.N., Rangari, V., Singh, B. and Chandan, B.K., 1992. Structure and hepatoprotective activity of a biflavonoid from *Canarium manii*. *Planta Med.* 58(6): 493- 495.
- Anandan, R. and Devaki, T., 1999. Hepatoprotective effect of *Picrorrhiza kurroa* on tissue defence system in dgalactosamine- induced hepatitis in rats. *Fitoterapia* 70(1): 54-57.
- Aniya, Y., Koyama, T., Miyagi, C., Miyahira, M., Inomata, C., Kinoshita, S. and Ichiba, T., 2005. Free radical scavenging and hepatoprotective actions of the medicinal herb, *Crassocephalum crepidioides* from the Okinawa Islands. *Biol. Pharm. Bull.* 28(1): 19-23.
- Asha, V.V., Sheeba, M.S., Suresh, V. and Wills, P.J., 2007. Hepatoprotection of *Phyllanthus maderaspatensis* against experimentally induced liver injury in rats. *Fitoterapia* 78(2): 134-141.
- Babu, B.H., Shylesh, B.S. and Padikkala, J., 2001. Antioxidant and hepatoprotective effect of *Acanthus ilicifolius*. *Fitoterapia* 72(3): 272-277(6).
- Banskota, A.H., Tezuka, Y., Adnyana, I.K., Xiong, Q., Hase, K., Tran, K.Q., Tanaka, K., Saiki, I. and Kadota, S., 2003. Hepatoprotective effect of *Combretum quadrangulare* and its constituents. *Biol. Pharm. Bull.* 23(4): 456-460.
- Bhakta, T., Mukherjee, P.K., Mukherjee, K., Banerjee, S., Mandal, S.C., Maity, T.K., Pal, M. and Saha, B.P., 1999. Evaluation of hepatoprotective activity of *Cassia fistula* leaf extract. *J. Ethnopharmacol.* 66(3): 277-282.
- Bhandarkar, M.R. and Khan, A., 2004. Antihepatotoxic effect of *Nymphaea stellata* Willd., against carbon tetrachloride induced hepatic damage in albino rats. *J. Ethnopharmacol.* 91(1): 61-64.
- Bishayee, A., Sarkar, A. and Chatterjee, M., 1995. Hepatoprotective activity of carrot (*Daucus carota* L.) against carbon tetrachloride intoxication in mouse liver. *J. Ethnopharmacol.* 47(2): 69-74(6).
- Bodakhe, S.H. and Ram, A., 2007. Hepatoprotective Properties of *Bauhinia variegata* Bark Extract. *Yakugaku Zasshi.* 127: 1503- 1507.

- Chander, R., Srivastava, V., Tandon, J.S. and Kapoor, N.K., 1995. Antihepatotoxic activity of diterpenes of *Andrographis paniculata* (Kalmegh) against Plasmodium berghei induced hepatic damage in Mastomys natalensis. *Int. J. Pharmacogn.* 33(2): 135-138.
- Chih, H.W., Lin, C.C. and Tang, K.S., 1996. The hepatoprotective effects of Taiwan folk medicine Ham-hong-chho in rats. *Am. J. Chinese Med.* 24: 231-240.
- Daba, M.H. and Abdel-Rahman, M.S., 1998. Hepatoprotective activity of thymoquinone in isolated rat hepatocytes. *Toxicol. Lett.* 95(1): 23-29.
- Dahiru, D., William, E.T. and Nadro, M.S., 2005. Protective effect of *Ziziphus mauritiana* leaf extract on carbon tetrachloride-induced liver injury. *Afr. J. Biotechnol.* 4(10): 1177-1179.
- Das, B.K., Bepary, S., Datta, B.K., Chowdhury, A.A., Ali, M.S. and Rouf, A.S., 2008. Hepatoprotective activity of *Phyllanthus reticulate*. *Pak. J. Pharm. Sci.* 21(4): 333-337.
- Dhanabal, S.P., Syamala, G., Satish Kumar, M.N. and Suresh, B., 2006. Hepatoprotective activity of the Indian medicinal plant *Polygala arvensis* on d-galactosamine-induced hepatic injury in rats. *Fitoterapia* 77(6): 472-474.
- Diallo, B., Vanhaelen-Fastre, R., Vanhaelen, M., Fiegel, C., Joyeux, M., Roland, A. and Fleurentin, J., 1992. Further studies on the hepatoprotective effects of *Cochlospermum tinctorium* rhizomes. *J. Ethnopharmacol.* 36(2): 137-142.
- Du, X.M., Sun, N.Y., Chen, Y., Irino, N. and Shoyama, Y., 2000. Hepatoprotective aliphatic glycosides from three *Goodyera* species. *Biol. Pharm. Bull.* 23(6): 731-734.
- Fakurazi, S., Hairuszah, I. and Nanthini, U., 2008. *Moringa oleifera* Lam. prevents acetaminophen induced liver injury through restoration of glutathione level. *Food Chem. Toxicol.* 46(8): 2611-2615.
- Fleurentin, J., Hoefler, C., Lexa, A., Mortier, F. and Pelt, J.M., 1986. Hepatoprotective properties of *Crepis rueppellii* and *Anisotes trisulcus*: two traditional medicinal plants of Yemen. *J. Ethnopharmacol.* 16(1): 105-111.
- Garg, H.S., Bhandari, S.P.S., Tripathi, S.C., Patnaik, G.K., Puri, A., Saxena, R. and Saxena, R.P., 1994. Antihepatotoxic and immunostimulant properties of iridoid glycosides of *Scrophularia koelzii*. *Phytother. Res.* 8(4): 224-228.
- Germano, M.P., Sanogo, R., Costa, C., Fulco, R., D'angelo, V., Torre, E.A., Viscomi, M.G. and De Pasquale, R., 1999. Hepatoprotective properties in the rat of *Mitracarpus scaber*. *J. Pharm. Pharmacol.* 51(6): 729-734.



- Gilani, A.H., Jabeen, Q., Ghayur, M.N., Janbaz, K.H. and Akhtar, M.S., 2005a. Studies on the antihypertensive, antispasmodic, bronchodilator and hepatoprotective activities of the *Carum copticum* seed extract. *J. Ethnopharmacol.* 98: 127-135.
- Gilani, A.H., Yaesh, S., Jamal, Q. and Ghayur, M., 2005b. Hepatoprotective activity of aqueous-methanol extract of *Artemisia vulgaris*. *Phytother. Res.* 19(2): 170-172.
- Gilani, A.H. and Janbaz, K.H., 1995a. Preventive and Curative Effects of *Artemisia absinthium* on acetaminophen and carbontetrachloride-induced hepatotoxicity. *Gen. Pharmacol.* 26(2): 309-315.
- Gilani, A.H. and Janbaz, K.H., 1995b. Studies on protective effect of *Cyperus scariosus* extract on acetaminophen and CCl<sub>4</sub>-Induced hepatotoxicity. *Gen. Pharmacol.* 26(3): 627-631.
- Gilani, A.H., Janbaz, K.H. and Shoaib, A.M., 1996. Selective protective effect of an extract from *Fumaria parviflora* on paracetamol-induced hepatotoxicity. *Gen. Pharmacol.* 27(6): 979-983.
- Gupta, M., Mazumder, K.U., Kumar, S.T., Gomathi, P. and Kumar, R.S., 2004. Antioxidant and Hepatoprotective Effects of *Bauhinia racemosa* against Paracetamol and Carbontetrachloride induced liver damage in rats. *IJPT.* 3: 12-20.
- Gupta, N.K. and Dixit, V.K., 2009. Evaluation of hepatoprotective activity of *Cleome viscosa* Linn. Extract. *Indian J. Pharmacol.* 41: 36-40.
- Gyamfi, M.A., Yonamine, M. and Aniya, Y., 1999. Free-radical scavenging action of medicinal herbs from Ghana: *Thonningia sanguinea* on experimentally-induced liver injuries. *Gen. Pharmacol.* 32(6): 661- 667.
- Hase, K., Kasimu, R., Basnet, P., Kadota, S. and Namba, T., 1997a. Preventive effect of lithospermate B from *Salvia miltorhiza* on experimental hepatitis induced by carbon tetrachloride or Dgalactosamine/ lipopolysaccharide. *Planta Med.* 63(1): 22-26.
- Hase, K., Li, J., Basnet, P., Xiong, Q., Takamura, S., Namba, T. and Kadota, S., 1997b. Hepatoprotective principles of *Swertia japonica* on Dgalactosamine/ lipopolysaccharide-induced liver injury in mice. *Chem. Pharm. Bull.* 45(11): 1823-1827.
- Hewawasam, R.P., Jayatilaka, K.A.P.W., Pathirana, C. and Mudduwa, L.K.B., 2004. Hepatoprotective effect of *Epaltes divaricata* extract on carbon tetrachloride-induced hepatotoxicity in mice. *Indian J. Med. Res.* 120: 30-34.

- Hyun, C., Jun, J.Y., Song, E.K., Kang, K.H., Baek, H.Y., Ko, Y.S. and Kim, Y.C., 2001. Bakuchiol: A hepatoprotective compound of *Psoralea corylifolia* on tacrine-induced cytotoxicity in Hep G2 cells. *Planta Med.* 67(8): 750- 751.
- Jadon, A., Bhadauria, M. and Shukla, S., 2007. Protective effect of *Terminalia bellerica* Roxb. and gallic acid against carbon tetrachloride-induced damage in albino rats. *J. Ethnopharmacol.* 109(2): 214-218.
- Jafri, M.A., Jalis, S.M., Javed, K. and Singh, S., 1999. Hepatoprotective activity of leaves of *Cassia occidentalis* against paracetamol and ethyl alcohol intoxication in rats. *J. Ethnopharmacol.* 66(3): 355-361.
- Jamshidzadeh Akram, Fereidooni Fatema, Salehi Zohreh and Niknahad, Hossein, 2005. Hepatoprotective activity of *Gundelia tourenfortii*, *J. Ethnopharmacology* 101: 233–237.
- Janakat, S. and Al-Merie, H., 2002. Evaluation of hepatoprotective effect of *Pistacia lentiscus*, *Phillyrea latifolia* and *Nicotiana glauca*. *J. Ethnopharmacol.* 83: 135-138.
- Janbaz, K.H., Saeed, S.A. and Gilani, A.H., 2002. Protective effect of rutin on paracetamol and CCl4-Induced hepatotoxicity in rodents. *Fitoterapia* 73: 557-563.
- Kadarian, C., Broussalis, A.M., Mino, J., Lopez, P., Gorzalczany, S., Ferraro, G. and Acevedo, C., 2002. Hepatoprotective activity of *Achyrocline satureioides* (Lam) D.C. *Pharmacol. Res.* 45(1): 57-61.
- Kim, E.Y., Kim, E.K., Lee, H.S., Sohn, Y., Soh, Y., Jung, H.S. and Sohn, N.W., 2007a. Protective effects of *Cuscutae semen* against Dimethylnitrosamine-Induced Acute Liver Injury in Sprague-Dawley Rats. *Biol. Pharm. Bull.* 30(8): 1427-1431.
- Kim, K.H., Kim, Y.H. and Lee, K.R., 2007b. Isolation of quinic acid derivatives and flavonoids from the aerial parts of *Lactuca indica* L. and their hepatoprotective activity *in vitro*. *Bioorg. Med. Chem. Lett.* 17(24): 6739-6743.
- Kinoshita, S., Inoue, Y., Nakama, S., Ichiba, T. and Aniya, Y., 2007. Antioxidant and hepatoprotective actions of medicinal herb, *Terminalia catappa* L. from Okinawa Island and its tannin corilagin. *Phytomed.* 14(11): 755-762.
- Krishna, M.G., Pallavi, E., Ravi, K.B., Ramesh, M. and Venkatesh, S., 2007. Hepatoprotective activity of *Ficus carica* Linn. leaf extract against carbon tetrachloride-induced hepatotoxicity in rats. *DARU* 15(3): 162- 166.
- Kumar, G., Sharmila, B.G., Vanitha, P.P., Sundararajan, M. and Rajasekara, P.M., 2004. Hepatoprotective activity of *Trianthema portulacastrum* L. against paracetamol and thioacetamide intoxication in albino rats. *J. Ethnopharmacol.* 92(1): 37-40.

- Kundu, R., Dasgupta, S., Biswas, A., Bhattacharya, A., Pal, B.C., Bandyopadhyay, D., Bhattacharya, S. and Bhattacharya, S., 2008. *Cajanus cajan* Linn (Leguminosae) prevents alcohol-induced rat liver damage and augments cytoprotective function. *J. Ethnopharmacol.* 118: 440- 447.
- Lin, C.C. and Huang, P.C., 2002. Antioxidant and hepatoprotective effects of *Acahopanax senticosus*. *Phytother. Res.* 14(7): 489-494.
- Lin, C.C., Lee, H.Y., Chang, C.H., Namba, T. and Hattori, M., 1996. Evaluation of the liver protective principles from the root of *Cudrania cochinchinensis* var. *gerontogea*. *Phytother. Res.* 10(1): 13-17.
- Lin, C.C., Lin, W.C., Chang, C.H. and Namba, T., 1995a. Antiinflammatory and hepatoprotective effects of *Ventilago leiocarpa*. *Phytother. Res.* 9(1): 11-15.
- Lin, C.C., Tsai, C.C. and Yen, M.H., 1995b. The evaluation of Hepatoprotective effects of Taiwan folk medicine "Teng – Khia – U". *J. Ethnopharmacol.* 45: 113-123.
- Lin, S.C., Lin, C.C., Lin, Y.H. and Shyuu, S.J., 1994. Hepatoprotective effects of Taiwan folk medicine: *Wedelia chinensis* on three hepatotoxininduced hepatotoxicity. *Am. J. Chin. Med.* 22(2): 155-168.
- Liu, C.T., Chuang, P.T., Wu, C.Y., Weng, Y.M., Wenlung, C and Tseng, C.Y., 2006. Antioxidative and in vitro hepatoprotective activity of *Bupleurum kaoi* leaf infusion. *Phytother. Res.* 20 (11): 1003-1008.
- Mandal, S.C., Saraswathi, B., Ashok Kumar, C.K., Mohana Lakshmi, S. and Maiti, B.C., 2000. Protective effect of leaf extract of *Ficus hispida* Linn. against paracetamol-induced hepatotoxicity in rats. *Phytother. Res.* 14(6): 457-459.
- Mankani, K.L., Krishna, V., Manjunatha, B.K., Vidya, S.M., Jagadeesh Singh, S.D., Manohara, Y.N., Raheman, A.U. and Avinash, K.R., 2005. Evaluation of hepatoprotective activity of stem bark of *Pterocarpus marsupium* Roxb. *Indian J. Pharmacol.* 37(3): 165-168.
- Manna, P., Sinha, M. and Sil, P., 2006. Aqueous extract of *Terminalia arjuna* prevents carbontetrachloride-induced hepatic and renal disorders. *BMC Complement. Altern. Med.* 6: 33.
- Mantena, S.K., Mutalik, S., Srinivasa, H., Subramanian, G.S., Prabhakar, R.K.R., Srinivasan, K.K. and Unnikrishnan, M.K., 2005. Antiallergic, Antipyretic, Hypoglycemic and Hepatoprotective Effects of Aqueous Extract of *Coronopus didymus* Linn. *Biol. Pharm. Bull.* 28(3): 468.
- Matsuda, H., Morikawa, T., Fengming, X., Ninomiya, K. and Yoshikawa, M., 2004. New isoflavones and pterocarpane with Hepatoprotective activity from the stems of *Erycibe expansa*. *Planta Med.* 70(12): 1201-1209.

- Meera, R., Devi, P., Kameswari, B., Madhumitha, B. and Merlin, N.J., 2009. *Indian J. Exp. Biol.* 47(7): 584-590.
- Mohammed, M.A., Marzouk, S.A, Moharram, F.A., El-Sayed, M.M. and Baiuomy, A.R., 2005. Phytochemical constituents and hepatoprotective activity of *Viburnum tinus*. *Phytochem.* 66(23): 2780-2786.
- Mondal, S.K., Chakraborty, G., Gupta, M. and Mazumder, U.K., 2005. Hepatoprotective activity of *Diospyros malabarica* bark in carbontetrachloride intoxicated rats. *Eur. Bull. Drug Res.* 13: 25-30.
- Morita, T., Jinno, K., Kawagishi, H., Arimoto, Y., Suganuma, H., Inakuma, T. and Sugiyama, K., 2003. Hepatoprotective effect of myristin from nutmeg (*Myristica fragrans*) on lipopolysaccharide/d-galactosamine-induced liver injury. *J. Agric. Food Chem.* 51: 1560-1565.
- Murugaian, P., Ramamurthy, V. and Karmegam, N., 2008. Hepatoprotective activity of *Wedelia calendulacea* L. against acute hepatotoxicity in rats. *Res. J. Agr. Biol. Sci.* 4(6): 685-687.
- Nguyen, N.T., Banskota, A., Tezuka, Y., Quan, L.T., Nobukawa, T., Kurashige, Y., Sasahara, M. and Kadota, S., 2004. Hepatoprotective effect of taxiresinol and (7'R)-7'-hydroxylariciresinol on d-galactosamine and lipopolysaccharide-induced liver injury in mice. *Planta Med.* 70(1): 29- 33.
- Oh, H., Kim, D.H., Cho, J.H. and Kim, Y.C., 2004. Hepatoprotective and free radical scavenging activities of phenolic petrosins and flavonoids isolated from *Equisetum arvense*. *J. Ethnopharmacol.* 95: 421-424.
- Oh, H., Kim, J.S., Song, E.K., Cho, H., Kim, D.H., Park, S.E., Lee, H.S. and Kim, Y.C., 2002. Sesquiterpenes with hepatoprotective activity from *Cnidium monnieri* on tacrine-induced cytotoxicity in Hep G2 cells. *Planta Med.* 68(8): 748-749.
- Olaleye, M.T., Adegboye, O.O. and Akindahunsi, A.A., 2006. *Alchornea cordifolia* extract protects wistar albino rats against acetaminophen-induced liver damage. *Afr. J. Biotechnol.* 5(24): 2439-2445.
- Oliveira, F.A., Chaves, M.H., Almeida, F.R.C., Lima, RCP, Silva, R.M., Maia, J.L., Brito, G.A.C., Santos, F.A. and Rao, V.S., 2005. Protective effect of  $\alpha$ - and  $\beta$ -amyrin, a triterpene mixture from *Protium heptaphyllum* (Aubl.) March. trunk wood resin, against acetaminophen-induced liver injury in mice. *J. Ethnopharmacol.* 98: 103-108.
- Opoku, A.R., Ndlovu, I.M., Terblanche, S.E. and Hutchings, A.H., 2007. *In vivo* hepatoprotective effects of *Rhoicissus tridentata* subsp. *cuneifolia*, a traditional Zulu medicinal plant against carbontetrachloride-induced acute liver injury in rats. *S. Afr. J. Bot.* 73(3): 372-377.

- Ozbek, H., Ugras, S., Dulger, H., Bayram, I., Tuncer, I., Ozturk, G. and Ozturk, A., 2003. Hepatoprotective effect of *Foeniculum vulgare* essential oil. *Fitoterapia* 74(3): 317-319.
- Park, E.J., Zhao, Y.Z., Young, H.K., Jung, J.L. and Dong, H.S., 2004. Acanthoic acid from *Acanthopanax koreanum* protects against liver injury induced by tert-butyl hydroperoxide or carbon tetrachloride *in vitro* and *in vivo*. *Planta Med.* 70(4): 321-327.
- Rafatullah, S., Mossa, J.S., Ageel, A.M., Al-Yahya, M.A. and Tariq, M., 1991. Hepatoprotective and Safety Evaluation Studies on Sarsaparilla. *Int. J. Pharmacogn.* 29(4): 296-301.
- Raju, K., Anbuganapathi, G., Gokulakrishnan, V., Raj Kapoor, B., Jayakar, B. and Manian, S., 2003. Effects of Dried Fruits *Solanum nigrum* Linn. against carbontetrachloride-Induced Hepatic Damage in Rats. *Biol. Pharm. Bull.* 26(11): 1618.
- Rana, A.C. and Avadhoot, Y., 1991. Hepatoprotective effects of *Andrographis paniculata* against carbontetrachloride-induced liver damage. *Arch. Pharm. Res.* 14: 93-95.
- Rao, G.M.M., Rao, C.V., Pushpangadan, P. and Shirwaikar, A., 2006. Hepatoprotective effects of rubiadin, a major constituent of *Rubia cordifolia* Linn. *J. Ethnopharmacol.* 103(3): 484-490.
- Rao, K.S. and Mishra, S.H., 1997. Hepatoprotective activity of the whole plants of *Fumaria indica*. *Indian J. Pharm. Sci.* 59(4): 165-70.
- Rao, K.S. and Mishra, S.H., 1998. Antihepatotoxic activity of monomethyl fumarate isolated from *Fumaria indica*. *J. Ethnopharmacol.* 60(3): 207-213.
- Rawat, K.S., Mehrotra, A.S.A., Tripathi, S.C. and Shome, B.U., 1997. Hepatoprotective Activity of *Boerhaavia diffusa* L. Roots - A Popular Indian Ethnomedicine. *J. Ethnopharmacol.* 56, 61- 66.
- Ray, D., Sharatchandra, K. and Thokchom, I.S., 2006. Antipyretic, antidiarrhoeal, hypoglycemic and hepatoprotective activities of ethyl acetate extract of *Acacia catechu* Willd in albino rats. *Indian J. Pharmacol.* 38(6): 408-413.
- Sabir, S.M. and Rocha, J.B.T., 2008. Water-extractable phytochemicals from *Phyllanthus niruri* exhibit distinct *in vitro* antioxidant and *in vivo* hepatoprotective activity against paracetamol-induced liver damage in mice. *Food Chem.* 111(4): 845-851.
- Sadasivan, S., Latha, P.G., Sasikumar, J.M., Rajashekar, S., Shyamal, S. and Shine, V.J., 2006. Hepatoprotective studies on *Hedyotis corymbosa* (L) Lam. *J. Ethnopharmacol.* 106(2): 245-249.

- Sambath, K.R., Sivakumar, T., Sivarkumar, P., Nethaji, R., Vijayabasker, M., Perumal, P., Malaya, G. and Upal, K.M., 2005. Hepatoprotective and *in vivo* antioxidant effects of *Careya arborea* against carbontetrachlorideinduced liver damage in rats. *Intl. J. Mol. Med. Adv. Sci.* 1(4): 418- 424.
- Senthilkumar, K.T.M., Raj Kapoor, B. and Kavimani, S., 2005. Protective effect of *Enicostemma littorale* against carbontetrachloride-induced hepatic damage in rats. *Pharm. Biol.* 43(5): 485-487.
- Sethuraman, M.G., Lalitha, K.G. and Kapoor, B.R., 2003. Hepatoprotective activity of *Sarcostemma brevistigma* against carbon tetrachlorideinduced hepatic damage in rats. *Curr. Sci.* 84(9): 1186-1187.
- Shahjahan, M., Sabitha, K.E., Mallika, J. and Shyamala-Devi, C.S., 2004. Effect of *Solanum trilobatum* against carbon tetrachloride-induced hepatic damage in albino rats. *Indian J. Med. Res.*, 120: 194-198.
- Shim, S.B., Kim, N.J. and Kim, D.H., 2000.  $\beta$ -Glucuronidase inhibitory activity and hepatoprotective effect of 18 $\beta$ -glycyrrhetic acid from the rhizomes of *Glycyrrhiza uralensis*. *Planta Med.* 66(1): 40-43.
- Singanan, V., Singanan, M. and Begum, H., 2007. The Hepatoprotective Effect of Bael Leaves (*Aegle marmalos*) in alcohol induced liver injury in albino rats. *Int. J. Sci. Technol.*, 2(2): 83-92.
- Singh, A. and Handa, S.S., 1995. Hepatoprotective activity of *Apium graveolens* and *Hygrophila auriculata* against paracetamol and thioacetamide intoxication in rats. *J. Ethnopharmacol.* 49(3): 119- 126.
- Singh, B., Saxena, A.K., Chandan, B.K., Bhardwaj, V., Gupta, V.N., Suri, O.P. and Handa, S.S., 2001. Hepatoprotective activity of indigone: A bioactive fraction from *Indigofera tinctoria* Linn. *Phytother. Res.*, 15(4): 294- 297.
- Singh, B., Saxena, K., Chandan, B., Agarwal, S., Bhatia, M. and Anand, K., 1993. Hepatoprotective effects of ethanolic extract of *Eclipta alba* on experimental liver damage in rats and mice, *Phytother Res.* 7(2): 154-158.
- Song, E.K., Kim, J.H., Kim, J.S., Cho, H., Nan, J.X., Soku, D.H., Ko, G.J., Oh, H. and Ki, Y.C., 2003. Hepatoprotective phenolic constituents of *Rhodiola sachalinensis* on tacrine-induced cytotoxicity in Hep G2 cells. *Phytother. Res.* 17(5): 563-565.
- Suja, S.R., Latha, P.G., Pushpangadan, P. and Rajasekharan, S., 2003. Evaluation of hepatoprotective effects of *Rhinacanthus nasuta* root extracts. *J. Trop. Med. Plants* 4(2): 151-157.
- Suja, S.R., Latha, P.G., Pushpangadan, P. and Rajasekharan, S., 2004. Evaluation of hepatoprotective effects of *Helminthostachys zeylanica* (L.) Hook against carbon tetrachloride-induced liver damage in Wistar rats. *J. Ethnopharmacol.* 92: 61-66.

- Sumathy, T., Subramanian, S., Govindasamy, S., Balakrishna, K. and Veluchamy, G., 2001. Protective role of *Bacopa monniera* on morphine induced hepatotoxicity in rats. *Phytother. Res.* 15(7): 643- 645.
- Sung, S.H., Won, S.Y., Cho, N.J. and Gkim, C.Y., 1997. Hepatoprotective flavonol glycosides of *Saururus chinensis* herbs. *Phytother. Res.* 11(7): 500- 503.
- Tasduq, S.A., Kaiser, P., Gupta, D.K., Kapahi, B.K., Jyotsna, S., Maheshwari, H.S. and Johri, R.K., 2005. Protective effect of a 50% hydroalcoholic fruit extract of *Emblica officinalis* against anti-tuberculosis drugs induced liver toxicity. *Phytother. Res.* 19(3): 193-197.
- Tasduq, S.A., Singh, K., Satti, N.K., Gupta, D.K. and Suri, K.A., 2006. *Terminalia chebula* (fruit) prevents liver toxicity caused by sub-chronic administration of rifampicin, isoniazid and pyrazinamide in combination. *Hum. Exp. Toxicol.* 25(3): 111-118.
- Umadevi, S., Mohanta, G.P., Kalaiselvan, R., Manna, P.K., Manavalan, R., Sethupathi, S. and Shantha, K., 2004. Studies on hepatoprotective effect of *Flaveria trinervia*. *J. Nat. Rem.* 4(2): 168-173.
- Van, Puyvelde, L., Kayonga, A., Brioen, P., Costa, J., Ndimubakunzi, A., De Kimpe, N. and Schamp, N., 1989. The hepatoprotective principle of *Hypoestes triflora* leaves. *J. Ethnopharmacol.* 26(2): 121-127.
- Vijayan, P., Prashanth, H.C., Vijayaraj, P., Dhanaraj, S.A., Badami, S. and Suresh, B., 2003. Hepatoprotective effect of the total alkaloid fraction of *Solanum pseudocapsicum* leaves. *Pharm. Biol.* 41(6): 443-448.
- Vimal, V. and Devaki, T., 2004. Hepatoprotective effect of allicin on tissue defense system in galactosamine/endotoxin challenged rats. *J. Ethnopharmacol.* 90(1): 151-154.
- Visen, P.K.S., Shukla, B., Patnaik, G.K., Kaul, S., Kapoor, N.K. and Dhawan, B.N., 1991. Hepatoprotective activity of picroliv, the active principle of *Picrorhiza kurrooa*, on rat hepatocytes against paracetamol toxicity. *Drug Dev. Res.* 22(3): 209-219.
- Wang, M.Y., Anderson, G., Nowicki, D. and Jensen, J., 2008. Hepatic protection by *Morinda citrifolia* (noni) fruit juice against CCl<sub>4</sub>-induced chronic liver damage in female SD rats. *Plant Foods Hum. Nutr.* 63(3): 141- 145.
- Wu, J.B., Lin, W.L., Hsieh, C.C., Ho, H.Y., Tsay, H.S. and Lin, W.C., 2007. The hepatoprotective activity of kinsenoside from *Anoectochilus formosanus*. *Phytother. Res.* 21(1): 58-61.
- Wu, S.J., Wang, J.S., Lin, C.C. and Chang, C.H., 2001. Evaluation of hepatoprotective activity of Legumes. *Phytomedicine* 8(3): 213-219.

- Xiong, Q., Fan, W., Tezuka, Y., Ketut, A.I., Stampoulis, P., Hattori, M., Namba, T. and Kadota, S., 2000. Hepatoprotective effect of *Apocynum venetum* and its active constituents. *Planta Med.* 66(2): 127-133.
- Yadav, N.P. and Dixit, V.K., 2003. Hepatoprotective activity of leaves of *Kalanchoe pinnata* Pers. *J. Ethnopharmacol.* 86: 197-202.
- Yang, H., Sung, S.H. and Kim, Y.C., 2005. Two New Hepatoprotective Stilbene Glycosides from *Acer mono* leaves. *J. Nat. Prod.* 68(1): 101-103.
- Yoshikawa, M., Ninomiya, K., Shimoda, H., Nishida, N. and Matsuda, H., 2002. Hepatoprotective and Antioxidative Properties of *Salacia reticulata*: Preventive Effects of Phenolic Constituents on carbontetrachloride- Induced Liver Injury in Mice. *Biol. Pharm. Bull.* 25(1): 72.
- Yoshikawa, M., Xu, F., Morikawa, T., Ninomiya, K. and Matsuda, H., 2003. Anastatins A and B, new skeletal flavonoids with Hepatoprotective activities from the desert plant *Anastatica hierochuntica*. *Bioorg. Med. Chem. Lett.*, 13(6): 1045-1049.
- Zhang, Z.F., Fan, S.H., Zheng, Y.L., Lu, J., Wu, D.M., Shan, Q. and Hu, B., 2009. Troxerutin protects the mouse liver against oxidative stress-mediated injury induced by D-galactose. *J. Agric. Food Chem.* 57(17): 7731- 7736.

